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## SYNTHESIS OF A NEW $^{125}\text{I}$ -LABELED TYROSINE METHYL ESTER CONJUGATE OF ADENYLYL-(2'-5')-ADENYLYL-(2'-5')-[2',3'-DI-O-(2-CARBOXYETHYL)ETHYLIDENE]ADENOSINE

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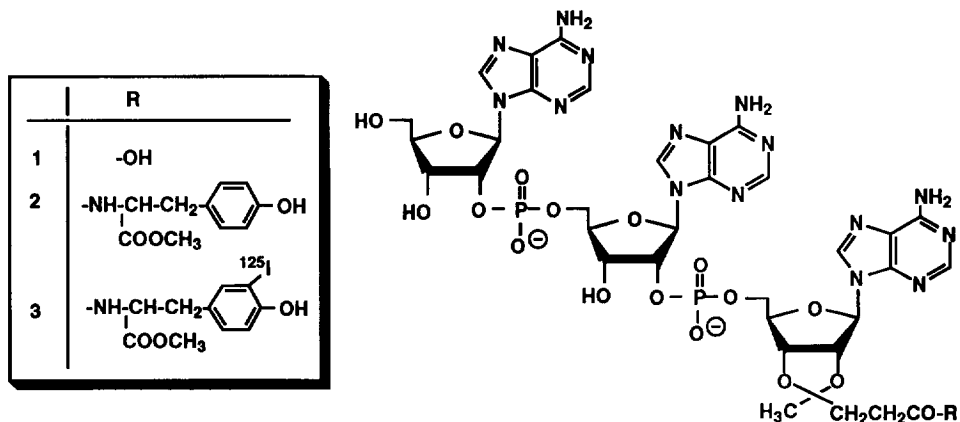
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**Abstract.** A novel  $^{125}\text{I}$ -labeled conjugate of 2',5'-oligoadenylate trimer for radioimmuno assay of 2',5'-oligoadenylates have been obtained by the reaction of adenylyl-(2'-5')-adenylyl-(2'-5')-[2',3'-di-O-(2-carboxyethyl)-ethylidene]adenosine (1) with tyrosine methyl ester followed by iodination of intermediate 2 with  $^{125}\text{I}$  in the presence of chloramine-T.  $^{125}\text{I}$ -labeled conjugate 3 was isolated by column chromatography on a TSK-gel HW-40 in 73% yield. Specific activity for the synthesized conjugate was approximately 1760 Ci/mmol.

### Introduction.

It is known that components of the 2-5A system are widely distributed in tissues and cells, and their activity varies with physiological conditions<sup>1</sup>. The functions of the 2-5A system outside the interferon treatment suggesting that it may be involved in the normal regulation of cellular RNA levels. However, the real importance and exact role of the 2-5A system is still unresolved. To examine its possible roles it is essential to have direct, sensitive assays for 2',5'-oligoadenylates, both 5'-phosphorylated and nonphosphorylated. The assays based on the inhibition of cell-protein synthesis through the activation of the 2-5A dependent RNase L<sup>2</sup>, or on the cytotoxic activity of nonphosphorylated derivatives<sup>3</sup>, are relatively insensitive and not suitable for the routine analysis of large numbers of samples<sup>4</sup>. The radioimmuno assay, using antibodies against 2',5'-oligoadenylates, is the most convenient method for this purpose. The major difficulty in the development of this assay lies in the synthesis of radioactive 2',5'-oligoadenylates of sufficiently high specific activity to give the required sensitivity. Some radioimmuno assays for 2',5'-oligoadenylates (5'-phosphorylated and nonphosphorylated) have been developed using [8- $^{14}\text{C}$ ]-A2'p5'A2'p5'A<sup>5</sup>, (A2'p)<sub>2</sub>A-[ $^{32}\text{P}$ ]pCp, and ppp(A2'p)<sub>2</sub>A-[ $^{32}\text{P}$ ]pCp<sup>4,6,7</sup>.  $^{125}\text{I}$ -labeled conjugates of alanyltyrosine methyl ester with periodate-oxidized 5'-triphosphates trimer and tetramer A2'p5'(A2'p)<sub>n</sub>5'A<sup>8,9</sup>, and  $^{125}\text{I}$ -labeled A2'p5'A succinyl tyrosine methyl ester<sup>10,11</sup>, as radiolabeled probes. Each of these compounds has a serious drawback which consists in low specific activity of [8- $^{14}\text{C}$ ]-A2'p5'A2'p5'A, high difference in the structure of periodate-oxidized and [ $^{32}\text{P}$ ]pCp containing derivatives with determined 2',5'-oligoadenylates, and insufficient stability of succinyl group of  $^{125}\text{I}$ -labeled A2'p5'A succinyl tyrosine methyl ester. To overcome these disadvantages we developed a new sensitive radioimmuno assay for 2',5'-oligoadenylates, using stable immuno-

genic and  $^{125}\text{I}$ -labeled conjugates attached to the adenylyl-(2'-5')-adenylyl-(2'-5')-[2',3'-di-O-(2-carboxyethyl)-ethylidene]adenosine (**1**) at the 2'-terminal carboxyl function. This type of conjugate allows to detect phosphorylated and nonphosphorylated 2',5'-oligoadenylates<sup>12,13</sup> at physiological concentrations (i.e. nanomolar range). Here we describe the synthesis of a new  $^{125}\text{I}$ -labeled tyrosine methyl ester derivative of 2',5'-A<sub>3</sub> trimer.



### Chemical Syntheses

**Adenylyl-(2'-5')-adenylyl-(2'-5')-[2',3'-di-O-(2-carboxyethyl)ethylidene]adenosine tyrosine methyl ester conjugate (**2**)**<sup>18</sup>. To a solution of bis-triethylammonium salt of trimer **1**<sup>14</sup> (6.3 mg, 4.7  $\mu\text{mole}$ ) in a mixture of DMF (300  $\mu\text{l}$ ) and  $\text{NEt}_3$  (8.3  $\mu\text{l}$ , 6.03 mg, 60  $\mu\text{mole}$ ), butyl chloroformate (7.8  $\mu\text{l}$ , 8.21 mg, 60  $\mu\text{mole}$ ) was added at 0°C, and the reaction mixture was stirred at room temperature for 40 min. Then a suspension of tyrosine methyl ester hydrochloride (6.96 mg, 30  $\mu\text{mole}$ ) and  $\text{NEt}_3$  (4.17  $\mu\text{l}$ , 3.03 mg, 30  $\mu\text{mole}$ ) in DMF (300  $\mu\text{l}$ ) was added, and the reaction mixture incubated at room temperature for 15 h. To the resulting homogeneous mixture, a saturated solution of ammonia in methanol (100  $\mu\text{l}$ ) was added and the mixture evaporated. The residue was dissolved in water (2 ml) and applied onto a DEAE-cellulose SS-23 column (2x16 cm;  $\text{HCO}_3^-$ -form). The product was eluted with a linear gradient of aqueous (0.01/0.3 M) TEAB buffer solution. The product containing fractions were collected and lyophilized to give a residue (2.6 mg; 36%) of conjugate **2**. UV<sup>17</sup> ( $\text{H}_2\text{O}$ ):  $\lambda_{\text{max}}$  260 nm,  $\epsilon = 36900$ .  $^1\text{H-NMR}^{17}$  ( $\text{D}_2\text{O}$ ): 8.17 (s, 1H), 8.11 (s, 1H), 7.91 (s, 2H), 7.83 (s, 1H) and 7.75 (s, 1H) (2-H, 8-H); 7.01 (d, 2H, o to PhOH,  $J=9.6$  Hz); 6.68 (d, 2H, m to PhOH,  $J=9.6$  Hz); 6.10 (d, 1H, 1'-H,  $J_{1',2'}=4.8$  Hz); 5.94 (d, 1H, 1'-H,  $J_{1',2'}=3.0$  Hz); 5.84 (d, 1H, 1'-H,  $J_{1',2'}=1.2$  Hz); 5.05 (m, 2H, 2'-H); 3.83 (s, 3H,  $\text{OCH}_3$ ); 2.46 (m, 2H,  $\text{CH}_2\text{CH}_2\text{CONH}$ ); 2.13 (m, 2H,  $\text{CH}_2\text{CH}_2\text{CONH}$ ); 1.38 (s, 3H,  $\text{CCH}_3$ ). Anal. Calcd for  $\text{C}_{57}\text{H}_{84}\text{N}_{18}\text{O}_2\text{P}_2$ : C, 48.78; H, 6.03; N, 17.96. Found: C, 48.59; H, 6.01; N, 17.88.

**$^{125}\text{I}$ -labeled adenylyl-(2'-5')-adenylyl-(2'-5')-[2',3'-di-O-(2-carboxyethyl)ethylidene]-adenosine tyrosine methyl ester conjugate (3).** To a solution of bis-triethylammonium salt of conjugate 2 (1.4 mg, 1.0 mmol) in 15 ml of 0.1 M sodium phosphate buffer (pH 7.5), a solution of  $\text{Na}^{125}\text{I}$  (100 MBq) in 5 ml of 0.1 M sodium hydroxide, and chloramine-T (20 mg, 71  $\mu\text{mol}$ ) in 10 ml of sodium phosphate buffer were added. The reaction mixture was stirred at room temperature for 15 min, diluted with a solution of  $\text{Na}_2\text{S}_2\text{O}_5$  (20 mg, 120 mmol) in 10 ml of sodium phosphate buffer. The resulting solution was applied onto a TSK-gel HW-40 column (1x25 cm), and the product eluted with sodium phosphate buffer, containing 0.1% of bovine serum albumin. The product containing fractions were collected and lyophilized to give 73 MBq (73%) of  $^{125}\text{I}$ -labeled conjugate 3 with 98% of radiochemical purity and a specific radioactivity of 1760 Ci/mmol.

## Results and Discussion

For the synthesis of radioactive labeled probe 3 trimeric adenylyl-(2'-5')-adenylyl-(2'-5')-[2',3'-di-O-(2-carboxyethyl)ethylidene]adenosine (**1**) was used as a starting compound<sup>14</sup> which implicates some structural advantages in this approach. Firstly, the synthesis of 2',3'-di-O-(2-ethoxycarbonyl)ethylidene-N<sup>6</sup>-benzoyl-adenosine - a key intermediate for the preparation of trimer **1** - resulted in formation of only one diastereomer with the unequivocally defined structure of the *endo* conformation of the methyl group, and the *exo* conformation of the carboxyethyl side chain<sup>15</sup>. Secondly, this modification keeps the ribofuranose ring at the 2'-terminal nucleoside intact and does not influence the spatial arrangement of the oligomer **1** vs. trimer 2'-5'A<sub>3</sub>. This conclusion follows from the identity of the ORD-spectra of both oligomers (data not shown). Thirdly, we have found that compound **1** does not undergo remarkable degradation upon incubation with rabbit serum at 37°C for 24 h.

The carboxyl function of trimer **1** was activated by treatment with butyl chloroformate to the intermediary mixed anhydride which on treatment with L-tyrosine methyl ester gave the desired conjugate **2** in 36% isolated yield after anion-exchange column chromatography. The structure of this compound was confirmed by UV and <sup>1</sup>H-NMR spectroscopy as well as elemental analysis.

Trimer **2** was iodinated with  $^{125}\text{I}$  by conventional chloramine-T method<sup>16</sup> to give  $^{125}\text{I}$ -tyrosine methyl ester conjugate of adenylyl-(2'-5')-adenylyl-(2'-5')-[2',3'-di-O-(2-carboxyethyl)ethylidene]adenosine (**3**), isolated in 73% yield by column chromatography on TSK-gel HW-40. The specific radioactivity of the  $^{125}\text{I}$ -labeled compound was 1760 Ci/mmol which suggests that nearly all the molecules of trimer **3** were iodinated with  $^{125}\text{I}$ . This novel conjugate was stable upon storage in a freezer for 2 months. Conjugate **3** was used as a labeled probe of which 90% was bound to an excess of antibodies and fully displaceable by trimer **2**.

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## References and Notes

1. Torrence, P.F. in *Biological response modifiers. New approach to disease intervention*. Torrence, P.F., Ed., Academic Press, Orlando; New York, **1985**, 77.
2. Williams B.R.G., Brown, R.E., Gilbert, C.S., Golgher, R.R., Wrechner, D.H., Roberts, W.K., Silverman, R.H., Kerr, I.M. *Methods Enzymol.* (S. Pestka, Ed.), **1981**, 79 B, 198.
3. Kimchi, A., Shure, H., Revel, M. *Nature (London)*, **1979**, 282, 849.
4. Knight, M., Wrechner, D.H., Silverman, R.H., Kerr, I.M. *Methods Enzymol.* (S. Pestka, Ed.), **1991**, 79 B, 216.
5. Sawai, H., Shinomiya, T. *J. Biochem.* **1982**, 92, 1723.
6. Knight, M., Cayley, P.J., Silverman, R.H., Wrechner, D.H., Gilbert, C.S., Brown, R.E., Kerr, I.M. *Nature (London)*, **1980**, 288, 189.
7. Hersh, S.L., Reid, T.R., Friedman, R., Stark, G.R. *J. Biol. Chem.* **1984**, 259, 1727.
8. Sawai, H., Ishibashi, K., Itoh, M., Watanabe, S. *J. Biochem. (Tokyo)*, **1985**, 98, 999.
9. Sawai, H., Haira, H., Ishibashi, K., Itoh, M. *J. Biochem. (Tokyo)*, **1987**, 101, 339.
10. Cailla, H., LeBorgne DeKaouel, C., Roux, D., Delage, M., Marti, J. *Proc. Natl. Acad. Sci. U.S.A.* **1982**, 79, 4742.
11. Lawrence, L., Roux, D., Marti, J., Cailla, H. *Molecul. Immunol.* **1987**, 24, 1033.
12. Kvasyuk, E.I., Kulak, T.I., Shulyakovskaya, S.M., Makarenko, M.V., Mikhailopulo, I.A. *Int. J. Purine and Pyrimidine Res.* **1991**, 2, 73.
13. Mikhailopulo, I.A., Kvasyuk, E.I., Kulak, T.I., Shulyakovskaya, S.M., Makarenko, M.V., Mikhailov S.N., Charubala, R., Pfeleiderer, W. *Nucleic Acids Res. Symp. Ser.* **1991**, 24, 67.
14. Kvasyuk, E.I., Kulak, T.I., Zaitseva, G.V., Mikhailopulo, I.A., Charubala, R., Pfeleiderer, W. *Tetrahedron Lett.* **1984**, 25, 3683.
15. Seela, F., Cramer, F. *Chem. Ber.* **1975**, 108, 1329.
16. Hunter, W.M. Greenwood, F.C. *Nature (London)*, **1962**, 194, 495.
17. <sup>1</sup>H-NMR spectrum was obtained on Bruker WM-360 spectrometer and is reported relative to tetramethylsilane ( $\delta$ -scale in ppm). Ultra-violet spectrum was recorded with Specord UV-VIS spectrophotometer. Elemental analysis was performed by the Microanalytical Laboratory at the Institute of Bioorganic Chemistry, Byelorussian Academy of Sciences in Minsk.
18. Butyl chloroformate was purchased from Fluka (Switzerland), tyrosine methyl ester hydrochloride from Reanal (Hungary), bovine serum albumin, DEAE-cellulose SS-23 from Serva, and TSK-gel HW-40 from Toyo Soda MFG, Co., Ltd. (Japan).

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